EVALUATION OF LACTICIN ADDITION ON GAS PRODUCTION KINETICS AND RUMEN METHANOGENESIS in Vitro

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ABSTRAK

Tujuan penelitian ini adalah untuk mengamati efek sebuah bakteriosin yang relatif baru yakni laktisin 3147 terhadap kinetika produksi gas dan emisi metana dalam lingkungan rumen secara in vitro. Penelitian terdiri dari dua buah eksperimen. Eksperimen 1 menggunakan rancangan acak lengkap (tiga ulangan) untuk mengamati efek penambahan laktisin (0 dan 10 µM) pada substrat berupa rumput kering dan campuran rumput:konsentrat (1:1) terhadap kinetika produksi gas dan emisi metana. Pada eksperimen 2, level penambahan laktisin ditingkatkan menjadi 0, 10, 25 dan 50 µM. Substrat, laktisin dan campuran cairan rumen-buffer diinkubasi pada suhu 39°C selama 24 jam. Produksi total gas dan emisi metana diamati selama dan setelah periode inkubasi. Hasil menunjukkan bahwa secara umum penambahan laktisin tidak berpengaruh secara nyata terhadap peubah produksi gas dibandingkan dengan kontrol, baik ketika ditambahkan pada substrat berupa rumput maupun rumput:konsentrat. Penambahan laktisin hingga konsentrasi 50 µM masih belum dapat menurunkan emisi metana, meskipun pada penambahan 25 µM terdapat kecenderungan menurunkan emisi metana. Substrat berupa rumput menghasilkan metana yang lebih rendah secara signifikan dibandingkan dengan rumput:konsentrat baik pada eksperimen 1 maupun 2 (P<0,05). Dapat disimpulkan bahwa penambahan laktisin hingga 50 µM masih belum dapat menurunkan emisi metana sehingga perlu diuji lebih lanjut pada konsentrasi yang lebih tinggi.

Kata kunci: laktisin, bakteriosin, metana, rumen, in vitro

ABSTRACT

The present study was aimed to investigate the effect of a novel bacteriocin, i.e. lacticin 3147, on gas production kinetics and methane emission under *in vitro* rumen environment in two consecutive experiments. In experiment 1, either no or 10 μ M of lacticin was added to hay or hay:concentrate (1:1, w/w) substrate. In experiment 2, the levels of lacticin additions were extended to 0, 10, 25 and 50 μ M. Samples were incubated in three replicates in both experiments at 39°C for 24 h. Total gas production and methane emission were measured during and after the incubation, respectively. Results revealed that, in general, addition of lacticin had limited significant effects on gas production parameters compared to control (without lacticin addition). Lacticin addition up to 50 μ M did not significantly decrease CH₄ emission, although a tendency of methane reduction existed when lacticin was added at 25 μ M. Hay diet produced significantly less methane emission than that of hay:concentrate diet both in experiment 1 and experiment 2 (P<0.05). It can be concluded that lacticin addition up to 50 μ M was unable to decrease CH₄ emission *in vitro* and therefore need to be tested further at higher concentrations.

Keywords: lacticin, bacteriocin, methane, rumen, in vitro

INTRODUCTION

Concern on global warming problem has received a lot of attention during the past decades.

A steady increase of earth surface temperature is considered to be due to the accumulation of some major green-house gases (GHG) such as carbon dioxide (CO_2), methane (CH_4), nitrogen oxide (N₂O) and chloro fluoro carbon (CFC) through anthropogenic activities. Ruminant production system is a source of CH₄ emission that contributes significantly by approximately 80 million tonnes of annual CH4 (Beauchemin et al., 2008). The gas itself is produced in the digestive tract of ruminants, particularly in the rumen, by methanogenic archaea via utilization of CO₂ and H₂ as their main substrates to form CH₄ (Morgavi et al., 2010). Although such reaction is a way for eliminating the toxicity of H_2 when being accumulated (McAllister and Newbold, 2008), the process can be optimized without confronting a normal rumen function. Further, in addition to its contribution to global warming, CH₄ emission is a form of energy losses from ruminants which may account up to 14% of their digestible energy intake (Cottle et al., 2011).

A number of nutritional strategies have been attempted to mitigate ruminal CH₄ emission. Bacteriocin, a proteinaceous substance produced by certain bacterial species to inhibit the growth of other species, is considered as a promising option in term of mitigating CH4. Nisin, a bacteriocin produced by certain strains of Lactococcus lactis, and bovicin HC5, a bacteriocin from Streptococcus bovis HC5, have been tested for their effects in inhibiting ruminal substances methanogenesis; both were demonstrated to mitigate ruminal CH₄ emission under in vitro incubation system and the magnitudes of the effects were dose-dependent (Callaway et al., 1997; Lee et al., 2002). Further investigation on other bacteriocins that are effective in mitigating ruminal CH₄ emissions has to be continued in order to obtain the most promising substance with regard to the objective while maintaining an optimum condition of general rumen fermentation.

Lacticin 3147 is a bacteriocin produced by Lactococcus lactis subsp. lactis DPC3147, a strain isolated from an Irish kefir grain that has a broad spectrum of inhibition and heat stable especially at low pH (Ryan *et al.*, 1996). Due to its antimicrobial properties particularly by inhibiting a wide range of Gram-positive bacteria, lacticin has been used in a number of food and biomedical applications such as in cheese making, for the treatment and/or prevention of mastitis in cattle, and for treating some antibiotic resistant human pathogens (Twomey *et al.*, 2002). Despite such beneficial effects of lacticin, the substance has not been assessed for its influence on rumen fermentation yet. Based on the anti-methanogenic effect of other bacteriocins, i.e. nisin and bovicin, therefore, lacticin may possess such property as well. The present study was therefore aimed to investigate the effect of lacticin addition on gas production kinetics and methane emission under *in vitro* rumen environment. The addition was done into two different substrates, i.e. hay or hay:concentrate (1:1, w/w) and at various levels of application.

MATERIALS AND METHODS

Experimental Design

Lacticin 3147 was obtained from Dairy Products Research Centre, Teagasc, Moorepark, Republic of Ireland. Evaluation of the respective bacteriocin as an anti-methanogenic agent was performed in two consecutive experiments. In Experiment 1, a factorial completely randomized design was applied to examine the effects of different levels of lacticin (0 and 10 µM) added to hay or hay:concentrate (1:1, w/w) substrates on gas production kinetics and CH₄ emission. Experiment 2 was conducted as a continuation of Experiment 1 in order to confirm the results obtained and to increase the levels of lacticin used. The experimental set up of Experiment 2 was almost similar to Experiment 1, except that the levels of lacticin were extended to 0, 10, 25 and 50 µM. In both experiments, samples were incubated in three replicates.

In Vitro Gas Production

Amounts of 380 mg hay or hay:concentrate substrates (containing approximately 93% dry matter) as basal diets were weighed and transferred into 100 ml calibrated glass syringes (Haeberle Labortechnik GmbH & Co. KG, Lonsee, Germany). The substrates were then added with 30 ml of incubation medium (consisted of 10 ml rumen liquor and 20 ml buffer) by following the method of Menke and Steingass (1988). The rumen fluid and particulate matter were collected before the morning feeding from two rumen fistulated cows fed on roughage and concentrate based diets, mixed, homogenized, strained and filtered through 100 µm nylon net. The glassware used was kept at approximately 39°C and flushed with CO₂ before use. Lacticin was prepared by solubilizing it in sodium phosphate buffer at pH 6.8. Subsequently, the substance was injected (≤1 ml) into 100 ml calibrated syringe from the syringe nozzle before dispensing buffered rumen liquor. The 30 ml

buffered medium containing rumen microbes was thereafter dispensed into the syringes, and the incubation was carried out at $39 \text{ }^{\circ}\text{C}$ for 24 h.

Chemical Analysis

Hay and concentrate used as substrates in the present experiments were subjected to proximate (AOAC, 1990) and Van Soest analyses (Van Soest et al., 1991). Dry matter (DM) was determined by drying the samples at 105°C for 16 h. Crude protein (CP) was analysed using Kjeldahl method by considering $CP = N \times 6.25$, whereas ether extract (EE) was analysed based on Soxhlet extraction system using petroleum ether as the solvent. For the cell wall fractions, neutral detergent fiber (NDF) and acid detergent fiber (ADF) values are expressed inclusive of residual ash and without addition of amylase. Non-fiber carbohydrate was obtained by subtracting CP, EE and NDF from organic matter. Chemical composition of the substrates is presented in Table 1.

Gas Reading and Methane Determination

During 24 h incubation period, the total gas was recorded at various time point interval from the calibrated scale on the syringe, i.e. at 0, 4, 8 and 24 h. At the end of incubation, methane production was measured using an infrared (0-30% range) methane analyser (Pronova Analysentechnik GmbH & Co. KG, Berlin, Germany) calibrated against 10.6% methane (Jayanegara et al., 2009a). Technically, after measuring the total gas volume, the tubing of the syringe outlet was inserted into the inlet of the methane analyser. The display on the methane analyser gives methane as percent of the total gas and this value was used for calculation of methane in total gas volume.

Data Analysis

Kinetics of gas production during incubation period was fitted into an exponential equation proposed by Orskov and McDonald (1979) as follow:

 $p = a + b (1 - e^{-ct})$

Dependent variable p is the cumulative gas production (ml) at t hour incubation, whereas a, b and c are the constants of the equation. The constants are interpreted as gas production from soluble feed fraction (a), gas production from insoluble but fermentable fraction (b), and rate of gas production (c). Thus a+b is the theoretical Table 1. Chemical Composition of Hay (H), Concentrate (C) and Hay:Concentrate (HC; 1:1, w/w) Used as Substrates in the *in vitro* Incubation (in % Dry Matter)

Н	С	HC	
%%			
93.6	93.0	93.3	
89.3	95.6	92.5	
10.7	4.4	7.5	
11.2	19.3	15.3	
2.0	1.6	1.8	
52.8	17.3	35.1	
32.6	5.8	19.2	
23.3	57.4	40.3	
	H 93.6 89.3 10.7 11.2 2.0 52.8 32.6 23.3	H C 93.6 93.0 89.3 95.6 10.7 4.4 11.2 19.3 2.0 1.6 52.8 17.3 32.6 5.8 23.3 57.4	

* Defined as: organic matter – (crude protein + ether extract + neutral detergent fiber)

maximum gas production during incubation at infinite t hour (the asymptotic value).

Data obtained were analysed using factorial analysis of variance (ANOVA). The factors were substrate (hay or hay:concentrate, 1:1 w/w) and level of lacticin addition. Allocation of factors or treatments into incubation units were based on a completely randomized design. When at least a factor showed significantly different at P<0.05, the analysis was continued using Tukey's test for a more specific treatment comparison. All statistical analyses were performed using STATISTICA software version 6.0.

RESULTS AND DISCUSSION

Gas Production Kinetics

In general, addition of lacticin had limited significant effects on gas production parameters compared to control (without lacticin addition). This was true for hay and hay:concentrate substrates, and was consistently observed both in Experiment 1 (Table 2) and Experiment 2 (Table 3). However, there were few exceptions: addition of 10 μ M lacticin to hay substrate decreased gas production at 4 and 8 h (P<0.05; Experiment 1), and addition of 50 μ M lacticin to hay:concentrate decreased gas production at 24 h (P<0.05; Experiment 2). The respective results suggest that

lacticin addition up to 50 μ M does not influence fermentation and digestion of carbohydrate (both fiber and starch). Since lacticin is basically a polypeptide, hence, addition of lacticin is theoretically increases protein content in the *in vitro* system. Apart from the fact that gas production from protein fermentation is relatively small as compared to that of carbohydrate (Getachew *et al.*, 1998), addition a proteinacous substance up to 50 μ M is considered to be too small to influence the change of total gas production.

The main source of variation that led to differences in gas production parameters was the different incubated substrates, i.e. between hay and hay:concentrate. It had to be noted that the differences did not appear during early fermentation period (<8 h), but the differences were significants at 24 h (P<0.05); higher gas production in the incubation of hay:concentrate compared to hay was observed both in experiment 1 and experiment 2 at 24 h fermentation period (P<0.05). For the gas production kinetics parameters, a+b was higher in the incubation of hay:concentrate compared to that of hay in experiment 1 and 2 (P<0.05). Interaction between substrate and lacticin addition was found to be significantly different in experiment 2; lower a+b was observed at higher levels of lacticin addition (P<0.05). Conversely to the previous parameter, c

was significantly higher in the incubation of hay compared to hay:concentrate both in experiment 1 and experiment 2 (P<0.05). No interaction between substrate and lacticin addition was observed for the rate of gas production parameter.

Higher gas production in the incubation of hay:concentrate substrate compared to hay was higher proportion of non-fiber due to carbohydrate and lower proportion of fiber fraction (both NDF and ADF) in hay:concentrate. It has been well-established that fiber, i.e. structural components of plant such as NDF, ADF, cellulose and lignin, is negatively influenced the feed quality and digestibility (Jayanegara et al., the contrary, 2009b). On non-structural carbohydrate like starch, which is presumably high in concentrate, is considered to be positively correlated with feed quality. It is therefore not surprising that incubation of hay:concentrate substrate produced higher gas production over the hay only substrate. In agreement with the results, Jayanegara et al. (2009c) observed that substrates with higher ADF contents produced less total gas production and lower organic matter digestibility as shown by the correlation coefficients between the respective parameters. Further, addition of concentrate may stimulate rumen microbial activity and population and, in turn, contributes to the overall fermentation and digestion process.

Table 2. Influence of Addition of Lacticin 3147 to Different Substrates on Gas Production Kinetics (Experiment 1)

Substrate	Lacticin	Gas Production (mL)			a + b (mI)	a (m I /h)
(S)	(L; µM)	4 h	8 h	24 h	- a+b (mL)	c (mL/n)
Н	0	23.5 ^b	42.0 ^b	76.6 ^a	91.7 ^a	0.076 ^b
	10	21.7 ^a	39.8 ^a	74.0 ^a	90.5 ^a	0.071 ^b
HC	0	22.8 ^b	42.2 ^b	89.1 ^b	128.2 ^b	0.050 ^a
	10	22.7 ^b	41.3 ^b	85.6 ^b	118.3 ^b	0.054 ^a
SEM		0.216	0.310	1.985	5.314	0.004
P-value						
S		0.049	ns	< 0.001	< 0.001	< 0.001
L		0.001	0.002	ns	ns	ns
S*L		0.004	ns	ns	ns	ns

H, hay; HC, hay:concentrate (1:1, w/w); SEM, standard error of the mean Superscripts with different letters are significantly different at P<0.05

Substrate	Lacticin	Gas production (mL)			.1 (T)	(T / 1)
(S)	(L; μM)	4 h	8 h	24 h	- a+b (mL)	c (mL/h)
Н	0	25.3	45.0	77.2 ^{ab}	88.0 ^a	0.088 ^{bc}
	10	26.3	46.0	77.3 ^{ab}	86.7 ^a	0.093 ^{bc}
	25	26.0	45.8	75.0 ^a	83.1 ^a	0.098 ^c
	50	24.8	44.3	77.8 ^b	90.2 ^a	0.083 ^b
HC	0	25.0	45.3	90.8 ^d	119.9 ^c	0.059 ^a
	10	25.5	45.8	90.0 ^{cd}	115.7 ^{bc}	0.063 ^a
	25	26.3	46.8	91.8 ^d	117.5 ^{bc}	0.064 ^a
	50	25.5	45.5	87.5 ^c	109.7 ^b	0.067 ^a
SEM		0.148	0.181	1.646	3.63	0.004
P-value						
S		ns	ns	< 0.001	< 0.001	< 0.001
L		ns	ns	ns	ns	ns
S*L		ns	ns	ns	0.013	ns

Table 3. Influence of Addition of Lacticin 3147 to Different Substrates on Gas Production Kinetics (Experiment 2)

H, hay; HC, hay:concentrate (1:1, w/w); SEM, standard error of the mean

Superscripts with different letters are significantly different at P<0.05

Ruminal Methane Emission

Lacticin up to 50 µM did not significantly influence CH4 emission when added to hay or hay:concentrate substrate. This was consistently observed in experiment 1 (10 µM lacticin; Figure 1) and experiment 2 (10-50 µM lacticin; Figure 2). However, a tendency of methane reduction existed when lacticin was added at 25 µM (experiment 2) especially in the hay substrate. It appears that lacticin up to 50 µM is still ineffective to be used for modifying rumen fermentation, including for mitigating CH₄ emission. As for other bacteriocins like nisin and bovicin in which their effects are dose-dependent (Callaway et al., 1997; Lee et al., 2002), it seems that lacticin at such concentration is still below a threshold level to significantly decrease CH₄ emission. This certainly opens the opportunity for further subsequent studies to test lacticin at higher levels of addition. Apart from that, Kalmokoff et al. (1996) stated that the effectifity of a certain bacteriocin in the rumen depends on its stability against ruminal proteolytic. Therefore it is probable that lacticin may loss part of its activity due to proteolytic mechanism occurring intensively in rumen environment.

As observed for gas production parameter, variation of CH₄ emission due to substrate different was also existed. In this study, hay:concentrate diet produced higher CH₄ than that of hay as consistently observed both in experiment 1 and experiment 2 (P<0.05). Such a result apparently contrasts to a commonly known concept that increasing proportion of concentrate instead of forage in diet would reduce methane emission from ruminants (Beauchemin et al., 2008). The concept is based on a theory that of concentrate favors propionate feeding production in the rumen over the acetate so that more hydrogen is being utilized rather than being produced. Furthermore, the concentrate creates unconducive rumen environment for the archaea methanogens by increasing rate of passage, lowering pH and eliminating some population of protozoa where part of the methanogens are symbiotically living together with the protozoa (Moss et al., 2000).

It has to be noted, however, that methane decrease by increasing level of concentrate in the diet is usually expressed in a relative unit, such as relative to gross energy intake or digestible energy intake or relative to unit of animal product (such



Figure 1. Ruminal Methane Emission on the Addition of Lacticin 3147 (0 and 10 μ M) to Different Substrates (Experiment 1). H, hay; HC, hay:concentrate (1:1, w/w). P-value: substrate, <0.001; lacticin, ns; substrate*lacticin, n



Figure 2. Ruminal Methane Emission on the Addition of Lacticin 3147 (0, 10, 25 and 50 μ M) to Different Substrates (Experiment 2). H, hay; HC, hay:concentrate (1:1, w/w). P-value: substrate, <0.001; lacticin, ns; substrate*lacticin, ns

as per unit of milk produced or body weight). When methane emission is expressed in an absolute unit, such l/day or mol/day, interestingly, the emission is higher at higher level of concentrate in the diet. For instance, in an *in vitro* study, Garcia-Martinez *et al.* (2005) showed that methane productions of high, medium and low forage diets were 701, 754 and 812 μ mol, which indicated that higher proportion of concentrate led to a higher absolute methane emissions. In agreement with the study, Lovett *et al.* (2003) found that absolute methane emissions (in L/day) were higher when finishing beef heifers were fed

with a diet containing 60% concentrate as compared to those of fed with a diet containing 35% concentrate. But, methane emission per unit of animal product (per kg of live weight gain and carcass gain; in a relative unit) was reduced by lower forage to concentrate ratio. A plausible explanation to such contrasting results is that, although there is a shift towards more propionate by increasing proportion of concentrate in the diet, the concentrate is clearly more digestible than that of forage. As a consequence, on the whole, hydrogen is produced more as well as the methane emission.

CONCLUSION

Lacticin addition up to 50 μ M was unable to significantly decrease CH₄ emission *in vitro*. It seems that lacticin at such concentration is still below a threshold level to create such an effect. Despite the fact, a tendency of methane reduction existed when lacticin was added at 25 μ M. Further studies are therefore required to test lacticin at higher concentrations in order to decrease methanogenesis at simultaneously improving rumen fermentation activity.

REFERENCES

- Association of Official Analytical Chemists (AOAC). 1990. Official Methods of Analysis. 15th ed. Association of Official Analytical Chemists, Arlington, VA, USA.
- Beauchemin, K.A., M. Kreuzer, F. O'Mara and T.A. McAllister. 2008. Nutritional management for enteric methane abatement: a review. Aust. J. Exp. Agric. 48:21-27
- Callaway, T.R., A.M.S. Carneiro De Melo and J.B. Russell. 1997. The effect of nisin and monensin on ruminal fermentations *in vitro*. Curr. Microbiol. 35:90-96
- Cottle, D.J., J.V. Nolan and S.G. Wiedemann. 2011. Ruminant enteric methane mitigation: a review. Anim. Prod. Sci. 51:491-514
- Garcia-Martinez, R., M.J. Ranilla, M.L. Tejido and M.D. Carro. 2005. Effects of disodium fumarate on *in vitro* rumen microbial growth, methane production and fermentation of diets differing in their forage:concentrate ratio. Brit. J. Nutr. 94:71-77
- Getachew, G., M. Blümmel, H.P.S. Makkar and K. Becker. 1998. *In vitro* gas measuring

techniques for assessment of nutritional quality of feeds: a review. Anim. Feed Sci. Technol. 72:261-281

- Jayanegara, A., N. Togtokhbayar, H.P.S. Makkar and K. Becker. 2009a. Tannins determined by various methods as predictors of methane production reduction potential of plants by an *in vitro* rumen fermentation system. Anim. Feed Sci. Technol. 150:230-237
- Jayanegara, A., H.P.S. Makkar and K. Becker. 2009b. The use of principal component analysis in identifying and integrating variables related to forage quality and methane production. J. Indon. Trop. Anim. Agric. 34:241-247
- Jayanegara, A., A. Sofyan, H.P.S. Makkar and K. Becker. 2009c. Gas production kinetics, organic matter digestibility and methane production *in vitro* in hay and straw diets supplemented by tannin-containing forages. Med. Pet. 32:120-129.
- Kalmokoff, M.L., F. Bartlett and R.M. Teather. 1996. Are ruminal bacteria armed with bacteriocins? J. Dairy Sci. 79:2297-2306
- Lee, S.S., J.T. Hsu, H.C. Mantovani and J.B. Russell. 2002. The effect of bovicin HC5, a bacteriocin from *Streptococcus bovis* HC5, on ruminal methane production *in vitro*. FEMS Microbiol. Lett. 217:51-55
- Lovett, D., S. Lovell, L. Stack, J. Callan, M. Finlay, J. Conolly and F.P. O'Mara. 2003. Effect of forage/concentrate ratio and dietary coconut oil level on methane output and performance of finishing beef heifers. Livest. Prod. Sci. 84:135-146.
- McAllister, T.A., and C.J. Newbold. 2008. Redirecting rumen fermentation to reduce

methanogenesis. Aust. J. Exp. Agric. 48:7-13.

- Menke, K.H., and H. Steingass. 1988. Estimation of the energetic feed value obtained from chemical analysis and *in vitro* gas production using rumen fluid. Anim. Res. Dev. 28:7-55
- Morgavi, D.P., E. Forano, C. Martin and C.J. Newbold. 2010. Microbial ecosystem and methanogenesis in ruminants. Animal 4:1024-1036
- Moss, A.R., J.P. Jouany and J. Newbold. 2000. Methane production by ruminants: its contribution to global warming. Ann. Zootech. 49:231-253
- Orskov, E.R., and I. McDonald. 1979. The estimation of protein degradability in the rumen from incubation measurements weighted according to rate of passage. J. Agric. Sci. 92:499-503
- Ryan, M.P., M.C. Rea, C. Hill and R.P. Ross. 1996. An application in cheddar cheese manufacture for a strain of *Lactococcus lactis* producing a novel broad-spectrum bacteriocin, lacticin 3147. Appl. Environ. Microbiol. 62:612-619
- Twomey, D., R.P. Ross, M. Ryan, B. Meaney and C. Hill. 2002. Lantibiotics produced by lactic acid bacteria: structure, function and applications. Antonie van Leeuwenhoek 82:165-185
- Van Soest, P.J., J.B. Robertson and B.A. Lewis. 1991. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. J. Dairy Sci. 74:3583-3597