



Heat Properties of Polylactic Acid Biocomposites after Addition of Plasticizers and Oil Palm Frond Microfiber

Wida Banar Kusumaningrum^{a,*}, Firda Aulya Syamani^a, Lisman Suryanegara^a

^a Biomaterials Research Center - Indonesian Institute of Sciences, Cibinong, Bogor, Indonesia

* Corresponding author: wida.banar@biomaterial.lipi.go.id

<https://doi.org/10.14710/jksa.23.8.295-304>



Article Info

Article history:

Received: 29th May 2020

Revised: 14th July 2020

Accepted: 28th July 2020

Online: 31st August 2020

Keywords:

non-isothermal
 crystallization kinetics;
 polyethylene glycol;
 polylactic acid; oil palm frond
 microfiber; triacetin

Abstract

Poly(lactic acid) (PLA) is a biopolymer that can replace thermoplastic polymers such as polypropylene (PP) in various applications due to strength, young modulus, biocompatibility, biodegradability, good clarity, oil resistance, and oxygen barrier ability. However, PLA has some drawbacks, including brittle, high glass transition temperature (T_g), and low degradation and crystallization rates. Therefore, modification is needed with the addition of nucleating agents and plasticizers to overcome these limitations of PLA. This research aims to study the effect of plasticizers and microfibril cellulose of oil palm frond (OPF) on thermal stability and to review the crystallization kinetics of PLA biocomposites. Polyethylene glycol and triacetin were used as plasticizers. Thermal analysis was performed using Thermal Gravimetry analysis (TGA) and Differential Scanning Calorimetry (DSC). The crystallization kinetics study was analyzed using a modified Avrami model under non-isothermal conditions. PLAP4000 has better thermal stability than PLAP200 and PLAG with T_{onset} and T_{max} values reaching 349.17°C and 374.68°C, respectively, which are close to pure PLA. All types of plasticizers influenced decreasing the T_g value in the range of 27–42%, whereas OPF microfiber addition contributes to a T_g reduction of 37–55%. Crystallization kinetic study was informed for heterogeneous and simultaneous nucleation mechanisms with an n value range of about 2–3 for PLAP4000 and PLAOP4000. The crystallization rate was multiplied 4–9-fold for PLAOP200 and 2–3-fold for PLAOP4000.

1. Introduction

The substitution of fossil-based polymers such as polypropylene and polyethylene with polymers that are easily biodegradable in the environment such as polylactic acid and polyhydroxy butyrate is becoming very important, amid issues of sustainability and a sustainable green industry movement. These fossil-based thermoplastic polymers are widely used in various applications, so their use and management must be controlled because they cannot be biodegradable in the environment. Poly(lactic acid) (PLA) is a biopolymer that has the potential to replace thermoplastic polymers such as polypropylene (PP) for several applications, because it has good strength and firmness, with 2 times the tensile strength and 2.5 times the young modulus, higher than PP [1]. It is also biocompatible, biodegradable, good transparency, good oil resistance, and oxygen barrier

ability [2, 3, 4, 5]. In connection with these properties, PLA can be applied to packaging and health materials, as well as automotive components [2, 3, 4]. Besides, the PLA manufacturing process has advantages in the heating process's effectiveness compared to other biopolymers and is more energy-efficient than fossil-sourced polymers [2]. PLA, which is synthesized by fermentation of molasses from biological sources such as sugarcane and corn is believed to be more economical and affordable [6]. However, through the fermentation process, the PLA obtained produced different molecular weights and crystal structures. PLA is polymorphic, which has three dimensions of the main crystal structure, which are α (orthorhombic), β (parallel orthorhombic), and γ (antiparallel helices arrangement) [7, 8, 9].

The polymorph nature of the PLA requires control during the heating and cooling process. It affects the

crystal structure formed, which has an impact on changing its mechanical and physical properties. PLA has several limitations, including brittleness, high glass transition temperature (T_g), which is close to room temperature ranging from 56–65°C, low rates of degradation and crystallization [2, 4, 8, 10]. Efforts are being made to improve the properties of PLA through intramolecular and intermolecular modifications. Effective modifications include the addition of nucleating agents, plasticizers, and a combination of both [2, 4, 6, 11].

Modification with plasticizers aims to increase the elasticity of PLA. The plasticizer mechanism in PLA is to insert plasticizers between the PLA molecular chains, thereby decreasing the rotational energy of the C–C bonds, which results in a decrease in T_g [12]. Some plasticizers can be used in PLA, both low and high molecular weights, including glycerol, triacetin, polyethylene glycol, lactic acid oligomers, and plasticized starch [6, 13]. The addition of plasticizers with low molecular weight, such as citrate esters, glycerol, and glucose monoesters, can increase (ductility) and impact resistance [14]. The ease of molecular chain mobility due to plasticization, significantly affects PLA's thermal characteristics, especially T_g and crystallization temperature (T_c). The addition of plasticizers, both low and high molecular weight, can reduce the T_g of PLA [6, 14, 15, 16, 17, 18]. Some studies state an increase in T_c , which shows a more regular crystal structure [19, 20]. Zubir *et al.* [11], Choi *et al.* [14] and Septevani and Bhakri [12] emphasized that plasticizers with low molecular weight have the potential to migrate, thereby reducing the elasticity of PLA over a while. Meanwhile, the plastic composition that is too high will reduce the mechanical properties of PLA. Another modification is the addition of a nucleating agent to accelerate the formation of nuclei, one of which is by strengthening natural fibers. Apart from being a nucleating agent, oil palm frond (OPF) microfibril can also be used as a reinforcing agent, thereby increasing the mechanical properties of PLA composites. Several studies stated that there was a decrease in T_g of PLA with the addition of microfibril by about 3% [21], 5% [22] to 15% [23] compared to pure PLA. The addition of 10% MFC (microfibril cellulose) was able to increase the crystallinity of PLA up to 32% and decrease the cold crystallization temperature (T_{cc}) [22, 24]. Apart from increased crystallinity, the addition of MCC (microcrystalline cellulose) from oil palm empty fruit bunches also improved the heat stability of PLA [25].

In thermoplastic polymers such as PLA, thermal analysis is indispensable because it affects the choice of production processes, handling, and storage. Modifications with the addition of nucleating agents and plasticizers will affect the thermal characteristics of PLA composites, primarily if the heating and cooling strongly influence the microstructure. In this research, a study of the effect of 3 types of plasticizers is glycerol triacetate (GTA) and polyethylene glycol (PEG) with low molecular weight, and PEG with high molecular weight. Also, as a nucleating agent, microfibrils from oil palm frond (OPF) pulp were added. The mixing process generally uses an

injection process, while in this study, the mixing was carried out by casting with a solvent casting. Therefore, it is necessary to do a thermal analysis because of additives and differences in processes. This study aimed to study the effect of plasticizer and OPF microfibril on thermal characteristics and a review of crystallization kinetics on PLA composites.

2. Methodology

This research consists of raw materials preparation, synthesis of PLA biocomposites, thermal analysis of PLA biocomposites, and a review of crystallization kinetics of PLA biocomposites.

2.1. Material

Oil palm frond fiber was obtained from the PTPN VIII Sukamaju plantation, West Java. Processing fiber into pulp through a sulfate (kraft) process was carried out at the Center for Pulp and Paper (BBKK) Bandung. PLA (polylactic acid) D3001 semicrystalline was obtained from Natureworks, USA, with a specific gravity of 1.24 g/cm³ and a melt flow rate of 22 g/10 min. The plasticizers used were GTA (glycerol triacetate) and PEG (polyethylene glycol) with low and high molecular weight. GTA pro analysis was obtained from Wako Pure Chemical Industries Ltd with a density of 1.157–1.165 g/cm³ and a molecular weight of 218.2 g/mol. The PEG used had a low molecular weight of 190–200 g/mol (PEG 200) with a density of 1.157–1.165 g/cm³, while the high molecular weight was 3500–4500 g/mol (PEG 4000) with a density of 1.2 g/cm³. The PEG 200 and PEG 4000 pro analyzes were obtained from Merck, Indonesia. The materials used for the pulp bleaching process were sodium chloride 25% solution in water, and glacial acetic acid 100% pro analysis obtained from Merck Indonesia. Meanwhile, for the PLA composite manufacturing process, 99% dichloromethane (DCM), 96% ethanol, and 90% acetone were used technically purchased from Frisconina.

2.2. Bleaching of oil palm frond pulp

OPF pulp with 10 g oven-dry weight was added 600 mL of distilled water. Sodium chloride and glacial acetic acid as delignification agents were added to the mixture, 16 mL and 0.8 mL, respectively. The bleaching process was carried out in a water bath at 80°C for 1 hour with periodic stirring every 15 minutes. The lighter colored pulp was then filtered and washed with distilled water until the pH is neutral, odorless, and clean. The bleached pulp yield was then referred to as OPF. The bleached OPF pulp's chemical composition was 4.53% lignin, 9.22% hemicellulose, and 86.25% cellulose.

2.3. Biocomposite Film Preparation

The PLA biocomposite manufacturing process went through several stages: raw material preparation, mixing, and molding. The solvent exchange process was carried out in 2 stages, first, using 96% ethanol stirred with pulp for 1 hour, followed by acetone stirring for 1 hour. PLA was dissolved with DCM and added 10% (w/w) of plasticizer then stirred for 60 minutes. As much as 10% (w/w) of OPF pulp was added to the PLA solution and plasticizer, then stirred again for 15 minutes. After the

mixing process, the solution was spread out on a teflon mat and dried at room temperature for 24 hours. PLA films were molded using a hot press at 170°C for 30 seconds at a pressure of 1 MPa. Film sheets were cut with 5 x 5 mm dimensions for testing thermal properties using DSC and TGA. The test samples were coded, as shown in Table 1.

Table 1. Sample code for PLA biocomposite test

Sample codes	Explanation
PLA	Pure PLA
PLAG	PLA + GTA 10%
PLAP200	PLA + PEG200 10%
PLAP4000	PLA + PEG4000 10%
PLAO	PLA + OPF 10%
PLAOG	PLA + GTA 10% + OPF 10%
PLAOP200	PLA + PEG200 10% + OPF 10%
PLAOP4000	PLA + PEG4000 10% + OPF 10%

2.4. Morphological analysis of OPF pulp fibers

OPF pulp fiber morphological analysis was carried out using Field Emission Scanning Electron Microscope (FE-SEM) - Quattro S Thermo Fisher Scientific. The OPF pulp fibers were placed in a step without coating, then analyzes were carried out at magnifications of 100, 500, and 1000 times by scanning electrons at 2 kV.

2.5. Thermal Properties Analysis

2.5.1. Thermal Gravimetry Analysis (TGA)

TGA analysis using the TGA 4000 Perkin Elmer. A test sample of 10 mg was placed in a ceramic crucible in a combustion chamber flowing with nitrogen gas. The test sample is heated from 25°C to 500°C with a heating rate of 10°C/min.

2.5.2. Differential Scanning Calorimetry (DSC)

The characterization of the thermal properties of the PLA biocomposite was carried out using the DSC 4000 Pyris 1 Perkin Elmer, which was equipped with a cooler. The results of the DSC analysis are the thermal parameters of polymer, including T_g (glass transition temperature), T_m (melting temperature), T_{cc} (cold crystallization temperature), T_c (crystallization temperature), melting enthalpy and crystallization enthalpy.

The DSC analysis stages included heating from -25°C to 250°C with a heating rate of 10°C/min. Then the samples were held at 250°C for 1 minute. Then cooled from 250°C to -25°C with a cooling rate of 10°C/min. Samples were held at -25°C for 1 minute. Then the samples were heated again from -25°C to 250°C with a heating rate of 10°C/min. The results obtained are used to determine the degree of crystallization of the biocomposite using equation 1.

$$Xc = \frac{\Delta H_m}{w \Delta H_m^0} \times 100\% \quad (1)$$

where w is the weight fraction of PLA in the composite, ΔH_m is the melting enthalpy of the biocomposite obtained at the second heating stage, while

ΔH_m^0 is the melting enthalpy for PLA which has a degree of crystallinity of 100% with a value of 93 W/g [26].

2.6. Overview of Non-Isothermal Crystallization Kinetics

The crystallization kinetics of the PLA system with the addition of plasticizer and OPF was carried out in non-isothermal conditions. This condition was chosen because, in real conditions, the biocomposite manufacturing process is carried out through a continuous process, including injection molding, blow molding, and extrusion. The crystallization kinetics analysis stage was heating from a temperature of 25°C to 200°C with a heating rate of 40°C/min. Then the samples were held at 200°C for 5 minutes, followed by cooling from 200°C to 0°C with set cooling rates of 5, 10, and 15°C/min. Kinetics analysis was carried out at the cooling (exothermic) process stage. The crystallization kinetics analysis approach used the Avrami model [27, 28]. Based on the assumption that total crystallization is linear with the energy released during the crystallization process, the relative crystallization degree ($X(t)$) can be described by equation 2.

$$X(t) = \frac{\int_0^t (\Delta H_c/dt) dt}{\int_0^\infty (\Delta H_c/dt) dt} \quad (2)$$

$X(t)$ is the volume fraction of the polymer that crystallizes at time t , ΔH_c is the crystallization enthalpy during the time interval dt (heat flow), t is the measured time during the crystallization process, ∞ is the time when crystallization ends. The crystallization kinetics parameters use the relative crystallization degree relationship with a constant from the Avrami model and are described by equation 3.

$$X(t) = 1 - \exp(-Kt^n) \quad (3)$$

K denotes the Avrami velocity constant, which includes the process of nucleation and crystal growth while n is the Avrami exponent, which shows the crystal geometry information. This constant can be obtained by the stratified logarithmic equation in equation 4.

$$\log[-\ln(1 - X(t))] = n \log t + \log K \quad (4)$$

The modification of Avrami model for non-isothermal process is at t which has a relationship with temperature and cooling rate as described in equation 5 [29], where T is the temperature at the time of crystallization t , T_0 is the temperature at the beginning of crystallization, while \emptyset is the velocity cooling down. Calculation of Avrami constants K and n by connecting the log graph $[-\ln(1 - X(t))]$ with $\log t$ so that the results K as the intercept and n as the slope.

$$t = \frac{T - T_0}{\emptyset} \quad (5)$$

3. Results and Discussion

3.1. Morphology of OPF pulp fiber

The bleached OPF pulp has long fiber morphology with a high aspect ratio, as shown in Figure 1 (a). Figures 1 (b) and 1 (c) show that the fiber diameter ranges from 10–30 μm . According to Zimmermann *et al.* [30], fibers

with a diameter of less than 50 μm with a millimeter-scale length are called microfibrils.

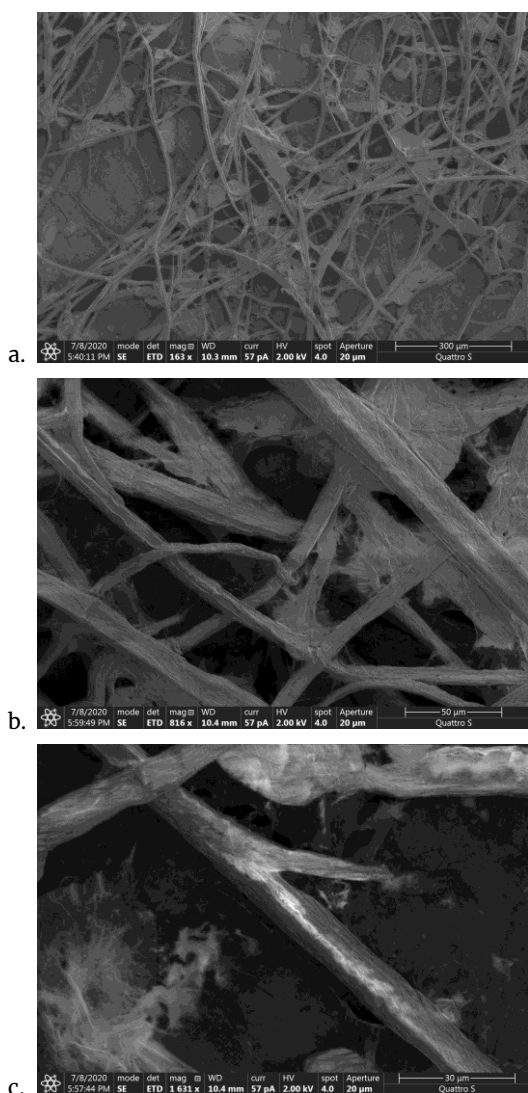


Figure 1. OPF pulp fiber morphology at magnifications of 100 times (a), 500 times (b), and 1000 times (c)

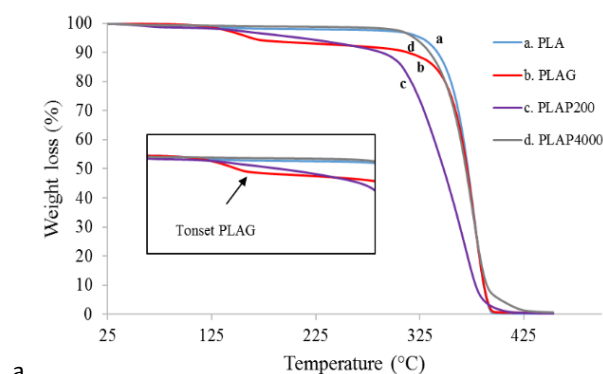
3.2. Characterization of thermal stability properties of PLA biocomposite through TGA analysis

The heat stability characteristic of PLA biocomposite is expressed by weight loss in temperature units obtained from TGA and DTG analysis. Figure 2(a) and Figure 2(b) show that the type of plasticizer affects the thermal stability of PLA biocomposites. The PEG-based plasticizer produces one decomposition peak, just like the pure PLA depicted in Figure 2 (b). However, there is a decrease in the initial decomposition temperature (T_{onset}) of PLAP200 and PLAP4000 compared to pure PLA, which are 11.5% and 2%, respectively. The lower molecular weight PEG produces PLA with lower thermal resistance, compared to the higher molecular weight PEG. This is because PEGs with higher molecular weight can be said more stable binds to the PLA polymer chain arrangement. Plasticizers with high molecular weight have longer polymer chains, so they are not easily decomposed [12]. Apart from that, it can also act as a protective layer to inhibit the permeability of volatile substances [31]. Quiles-Carrillo *et al.* [32] explained that the decrease in T_{onset} could be

caused by the evaporation of the plasticizer and the low molecular weight. On the other hand, the GTA plasticizer has two T_{onsets} , as shown in Figure 2 (a) and two decomposition peaks shown in Figure 2 (b). The PLAG T_{onset} is at 135.96°C and 353.33°C, while the peaks of decomposition are at 157.82°C and 376.69°C. These results may occur because GTA is not evenly dispersed in the PLA matrix, resulting in a separate decomposition of the plasticizer from the PLA matrix decomposition. Zubir *et al.* [11] explained that the incompatible phase difference between plasticizer and PLA resulted in two decomposition peaks. When viewed from the ease of dispersion or the effect of plasticizers on the mobility of the PLA polymer chain arrangement, PEG-based plasticizers are easier to disperse than GTA plasticizers.

The parameters of the thermal stability of PLA biocomposite obtained from the TGA and DTG analysis can be seen in Table 2, with observations only at the second peak for PLAG. The measured parameters in Table 2 obtained from the TGA thermogram include T_{onset} , initial decomposition temperature (T_0), decomposition peak temperature (T_{max}), and final decomposition temperature (T_f). Meanwhile, the DTG thermogram states the differential weight loss against temperature changes includes the temperature when the weight loss reaches 5% (T_5), 50% (T_{50}), and 95% (T_{95}), and char residue at 450°C. There was a decrease in T_{onset} , T_0 , T_{max} , T_f for all types of plasticizers. The reduction may be due to the plasticizer coating the surface and breaking down polymer interactions, particularly low molecular weight [12, 17]. PLAP200 has the most reduction compared to PLAG and PLAP4000. The observation on T_5 shows that PLAG decomposes faster than PLAP, then increases again after T_{50} and T_{95} . This indicates partial evaporation of GTA at the onset of heating [32].

In contrast, PLAP4000 provides better thermal resistance than PLAG and PLAP200 for all decomposition points (T_5 , T_{50} , T_{95}) and char residue, which shows a higher value than PLA, PLAG, and PLAP200. In this sense, PLAP has a broader range of processing, application, and storage temperatures than PLAG so that it can be processed and stored at temperatures above 200°C. The manufacturing process of PLA composites is in the temperature range of 170–180°C.



a.

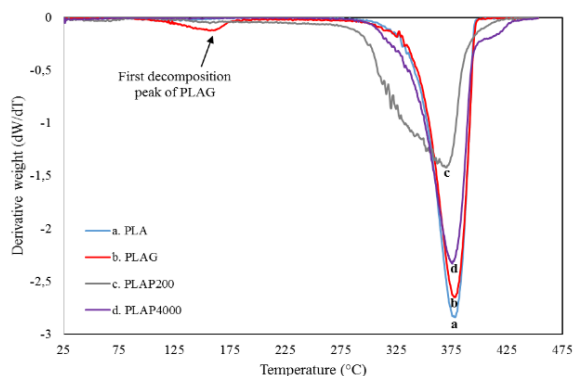
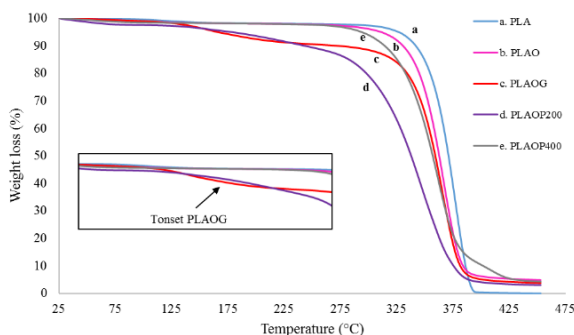
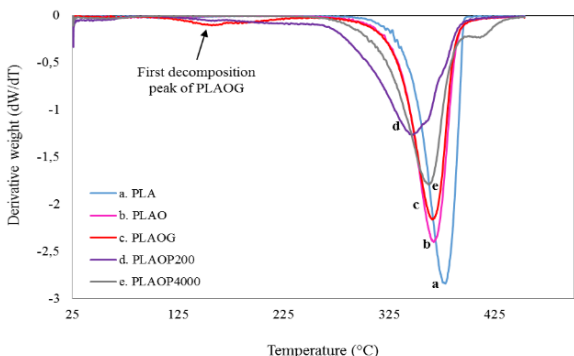


Figure 2. The PLA biocomposite thermogram with various types of plasticizers obtained through TGA (a) and DTG (b) analysis

The bleached OPF pulp in the form of microfibers is added to plasticized PLA composite with various types of plasticizers. The thermal stability characteristics of PLA composites with the addition of OPF and plasticizer obtained through TGA and DTG analysis are presented in Figures 3(a) and 3(b). The addition of OPF to PLA either with a plasticizer or not resulted in a decrease in T_{onset} . This is related to the thermal nature of lignocellulosic materials (hemicellulose and cellulose) which decompose in the range of 200–350°C [1, 33, 34]. However, this decrease can also be attributed to the low dispersed OPF microfibers in PLA despite the presence of plasticizers. These results differ from studies conducted by Haafiz *et al.* [25] and Safdari *et al.* [35], which showed an increase in thermal stability after adding micro and nanofibers to the PLA due to good dispersion, thus suppressing the mobility of the PLA chain. The same trend also occurs in the plasticized PLA/OPF system, where PLAOP4000 gives a higher T_{onset} value than PLAOP200, while PLAOG produces two T_{onsets} , as shown in Figure 3(a) and Figure 3(b).



a.



b.

Figure 3. Biocomposite PLA thermogram with various types of plasticizers and OPF obtained through TGA (a) and DTG (b) analysis

The weight-loss parameters for temperature are presented in Table 2, where the T_{onset} for PLAOG is only shown at the second peak. The DTG data review shows that T_0 , T_{max} , and T_t with the addition of OPF have decreased in value compared to pure PLA and plasticized PLA. A similar trend occurred in plasticized PLA composites with the addition of OPF for T5 and T50, where PLAOG had the lowest T5 value then increased when it reached T50 compared to PLAOP. However, when it reached T95, PLAOP4000 increased instantaneously with the char residue obtained reaching 0.43%. In this case, PEG4000 has the ability as a compatibilizer and plasticizer simultaneously [35]. Char residue of PLA composites with the addition of OPF, has a higher value than pure PLA and plasticized PLA. This shows that the carbon atoms in PLA are converted to gas, while the OPF microfibers become carbon.

Table 2. Value of thermal stability parameters based on the weight loss obtained through TGA and DTG analysis

Code	T_{onset} (°C)	T_0 (°C)	T_{max} (°C)	T_t (°C)	Weight loss			Char residue at 450°C (%)
					T5 (°C)	T50 (°C)	T95 (°C)	
PLA	356.44	345.31	377.4	395.1	326.24	370.97	389.08	0.001
PLAG	353.33	343.68	376.69	395.47	162.79	369.44	388.88	0.037
PLAP200	315.4	296.05	368	389.13	210.17	346.83	385.34	0.022
PLAP4000	349.17	336.65	374.68	395.78	319.99	369.01	402.14	0.063
PLAO	343.18	331.63	366.25	389.09	310.54	362.17	440.05	0.483
PLAOG	341.02	329.67	364.87	387.64	164.8	358.64	401.52	0.379
PLAOP200	309.43	291.44	346.39	385.35	176.87	337.41	389.13	0.300
PLAOP4000	330.14	318.22	361.17	386.7	293.44	356.74	429.79	0.431

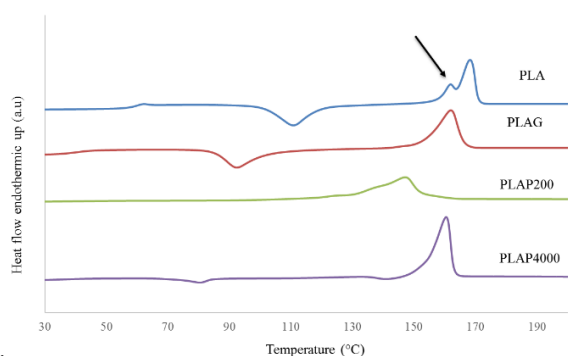
3.3. Thermal characteristics of PLA biocomposites through DSC analysis

DSC analysis provides information about the parameters of the thermal properties of PLA biocomposites, including T_m , T_c , and T_g , along with their energy. One of the weaknesses of PLA is that the high T_g value reaches 59.73°C, as presented in Table 3. The decrease in T_g value occurred with the addition of a plasticizer even up to 42.84% in the PLAP200. The peak of T_g widens, and there is a point shift for PLA with the addition of a plasticizer, as shown in Figure 4 (a). The decrease in T_g is due to the broader mobility of the plasticizer chain, making PLA more flexible [12, 17, 18, 35]. Semicrystalline PLA is unique. At a certain point during the endothermic process, it undergoes self-nucleating before melting occurs, which is known as cold crystallization, as shown in Figure 4 (a). A similar event occurred in PLAG, although the point shift occurred due to the decrease in T_g .

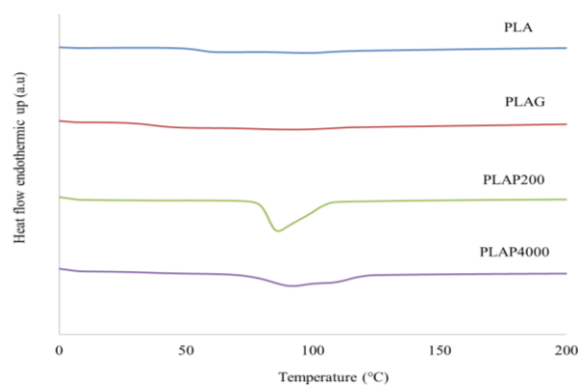
However, with the PEG plasticizer, there was a reduction in the area on the PLAP4000, as indicated by a decrease in the ΔH_{cc} value in Table 3. There was even no cold crystallization peak formation on the PLAP200. In this case, PEG can accelerate the crystal formation process in PLA. Cold crystallization formation indicate a slow crystallization process and amorphous PLA [23]. On

the other hand, the sloping ΔH_{cc} shows higher PLA crystallization, and the absence of T_{cc} indicates that PLA has crystallized in a certain order, which means that PEG functions as a crystal nucleating agent [20, 24]. The endothermic curve of pure PLA yielded two peaks, indicating that there was a melting of different crystal structures (regular and irregular) after cold crystallization (shown as arrows in Figure 4 (a).) This suggests that to obtain a regular crystal structure from irregular crystals requires a slower reorganization process [20, 22]. As explained by Salas-Papayanopolos *et al.* [18], there is a reorganization of unstable α' crystals into a more stable α . The addition of a plasticizer gave different results, where only one peak was formed, which referred to the formation of one type of crystal. These results also indicate that there is no crystal phase separation [17]. The melting point (T_m) values decrease considerably, reaching 31.8%; 42.8%; and 27.3% for PLAG, PLAP200, and PLAP4000, respectively, compared to pure PLA. This reduction in T_m contributes to the determination of the process production, application, distribution, and storage of PLA biocomposites in synergy with the decomposition temperature.

Figure 4 (b) shows that the PLAP200 and PLAP4000 produce exothermic curves with sharp peaks while PLA and PLAG are extensive. These results indicate that PLA and PLAG did not undergo a complete crystallization process. Therefore, PLA requires a nucleating agent to accelerate crystallization because PLA tends to undergo slow crystallization. The results presented in Figure 4 (b) and Table 3 show that PEG plasticizer can be a nucleating agent compared to GTA. The degree of crystallization (X_c) also increased for PLA with PEG plasticizers, both with low and high molecular weights. This may be because of the ease of insertion of molecules between the PLA polymer chains. Increasing chain mobility resulted in a decrease in T_g and T_{cc} , which contributed to an increase in the crystallization rate [17].



a.



b.

Figure 4. The DSC thermogram of PLA biocomposite with various types of plasticizers during endothermic (a) and exothermic (b) processes

Table 3. The value of the thermal parameters based on the DSC analysis

Code	T_g (°C)	T_m (°C)	ΔH_m (W/g)	T_{cc} (°C)	ΔH_{cc} (W/g)	T_c (°C)	ΔH_c (W/g)	X_c (%)
PLA	59.73	168.22	34.79	110.88	-26.41	63.8	-15.33	37.40
PLAG	40.72	161.91	34.75	92.53	-22.27	85.14	-19.13	37.36
PLAP200	34.14	147.11	43.11	-	-	86.52	-35.20	46.35
PLAP4000	43.45	166.52	44.20	86.67	-3.25	92	-25.50	47.53
PLAO	60.61	168.06	30.91	111.18	-20.43	97.82	-15.18	36.93
PLAOG	60.38	167.26	27.85	111.37	-19.37	98.47	-15.20	33.28
PLAOP200	26.56	149.95	29.62	-	-	95.66	-27.55	35.39
PLAOP4000	37.36	166.07	28.56	-	-	94.32	-22.35	34.12

The addition of OPF to the PLA matrix did not significantly affect the thermal properties of PLA biocomposites. Even there was an increase in T_g in PLAO and PLAOG, as presented in Table 3. In contrast to the PLAOP200 and PLAOP4000 biocomposites, the T_g value decreased by 55.5% and 37.5%. Meanwhile, similar characteristics to PLA occur in PLAO and PLAOG, where a cold crystallization occurs, and two peaks form on the endothermic curve (shown by arrows in Figure 5 (b)). Different results occurred in PLAOP200 and PLAOP4000 where no cold crystallization occurred, and one peak was formed on the endothermic curve. This is because OPF cannot accelerate the arrangement of the orderly crystal structure in PLAO and PLAOG. The addition of OPF to PLA and plasticized PLA caused a decrease in the degree of crystallization (X_c) compared to pure PLA. According to Safdari *et al.* [35], cellulose fibers to PLA can inhibit polymer chain mobility and crystal growth, thereby increasing stiffness, increasing in T_g value and decrease in X_c , if it is not adequately dispersed.

Figure 5 (b) shows the exothermic curve of the effect of OPF addition on PLA and plasticized PLA. Both PLAO and PLAOG show broad peaks like PLA, but for the PLAOP200 and PLAOP4000, the exothermic curve peaks are narrower. In this case, OPF can still act as nucleating agent for PLA with PEG plasticizers. PEG still contributes to the dispersion ability of OPF in the PLA matrix. Apart from being a plasticizer, PEG also functions as a compatibilizer, so that it has a better effect on the formation of PLA crystals compared to GTA. The crystallization peaks during the exothermic process are getting sharper, suggesting that there is a surface interaction between cellulose and PLA [23], thereby

increasing the nucleus-forming free area and encouraging crystallization [24].

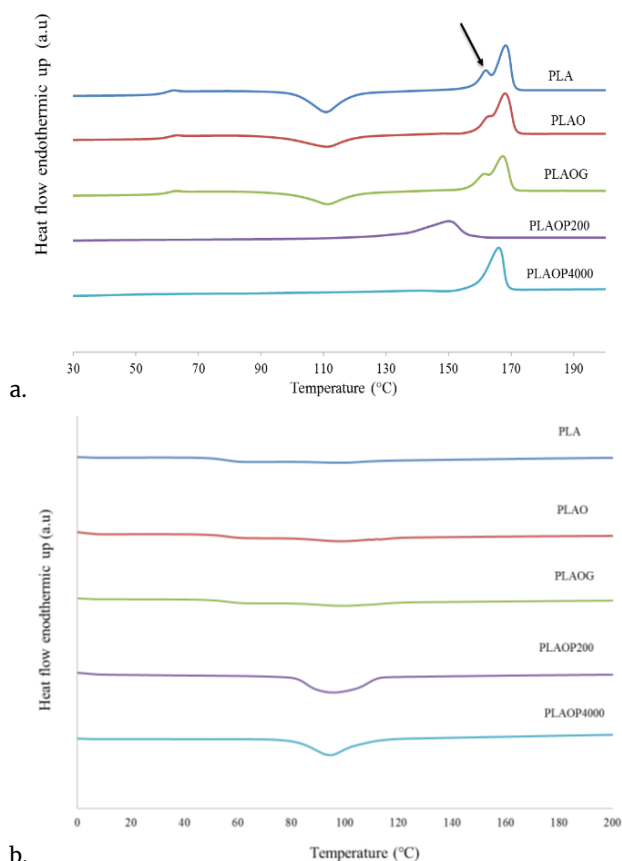


Figure 5. Thermogram analysis of PLA biocomposite DSC with various types of plasticizers and OPF during endothermic (a) and exothermic (b) processes

3.4. Overview of PLA Biocomposite Crystallization Kinetics

The crystallization kinetics of PLA biocomposites was carried out through the Avrami model approach, which explains the crystallization mechanism or the total solidification of thermoplastic polymers. Total crystallization includes the process of nucleation and crystal growth [27, 28]. The discussion on kinetics is more focused on PLA biocomposite with PEG plasticizer and the addition of OPF. The DSC analysis results show a much different change, especially in the crystallization process (exothermic conditions), compared to GTA plasticized PLA. The DSC analysis results showed that the PEG plasticizer and OPF microfibers were effective as nucleating agents. However, the effect can only be explained by the analysis of crystallization kinetics. Analysis of the crystallization kinetics using the Avrami model can determine the value of the constant K, which shows the total crystallization rate and the value of n, which shows the formed crystals geometry.

The crystallization process of PLA biocomposite through the Avrami model was obtained by connecting the stratified logarithmic graph of relative crystallization. The logarithmic of the temperature function in the non-isothermal crystallization process. The results of the graph plots are presented in Figure 6 for each cooling rate.

Figure 6 presents a linear line showing that the Avrami model fits a PLA biocomposite system at all cooling rates.

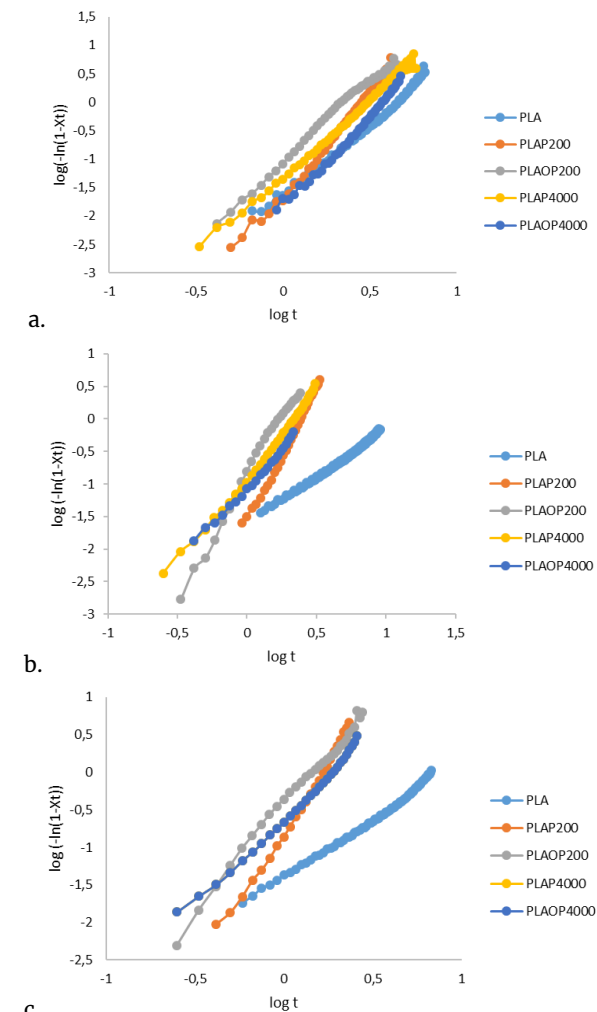


Figure 6. The relationship between $\log(-\ln(1-Xt))$ and $\log t$ of the Avrami model of PLA biocomposite at cooling rates of 5°C/minute (a), 10°C/minute (b), and 15°C/minute.

The crystallization rate constant (K) and crystal geometry are presented in Table 4. The K values indicated an increase for PEG 200 and PEG 4000 plasticized PLA at all cooling rates. This shows that the PEG plasticizer is effective as a nucleating agent and a crystal. However, with the addition of OPF, the PLAOP4000 only gave 3.7 and 2.3 times increase in cooling rates of 10°C/min and 15°C/min, respectively. In contrast to the PLAOP200, which still showed an increase of 4.2, 5.2, and 9.2 times for cooling rates of 5, 10, and 15°C/min, respectively. Different results were reported by Ding *et al.* [36], in which the presence of cellulose in PLA composites gave an increase in K value because cellulose was effective as a crystal nucleating agent.

The value of n is an integer number between 1- 4, where the value of n between 2-3 indicates the three-dimensional crystal growth dimension, while the n value <1 and n > 3 indicates 2-dimensional crystal [27, 28]. Besides showing the dimension of crystal growth, the value of n can describe the mechanism of crystal growth. The value of n <2 describes the mechanism of crystal

growth homogeneously and simultaneously. The values of $n = 2 - 3$ are heterogeneous and simultaneous, while more $n > 3$ are homogeneous and sporadic [4, 37, 38, 39]. Table 4 shows the results of n values between 2–3 on the PLAOP200, PLAP4000, and PLAOP4000. These results explain that the crystallization mechanism that occurs is heterogeneous crystallization and simultaneous crystallization with 3-dimensional crystals formed. Whereas in PLA, only with a speed of 5°C/min, which results in n at a value of 2–3. This shows that the crystallization of PLA is slow because, with a cooling rate of more than that, the value of n is 1–2. Different results were also obtained for the PLAP200, which resulted in a value of $n > 3$, which indicates that the crystals formed were denser. These results indicate that the crystallization process at a rate of 5–15°C/min is too slow for the PLAP200. Thus, PEG plasticizers can be said to be very effective as nucleating agents and increase the crystallization rate of PLA, which is generally between 1–3°C/min [8].

Table 4. Avrami Constants of PLA Biocomposites

Code	θ (°C/min)	K	n	R ²
PLA	5	0.02208	2.5547	0.990
	10	0.02336	1.4785	0.992
	15	0.03831	1.6037	0.991
PLAP200	5	0.02275	3.6256	0.994
	10	0.02574	4.1188	0.994
	15	0.16025	3.6982	0.993
PLAOP200	5	0.09389	2.7791	0.991
	10	0.12954	3.7247	0.990
	15	0.35197	2.7758	0.992
PLAP4000	5	0.05176	2.6357	0.995
	10	0.12639	2.6638	0.994
	15	0.24205	2.3143	0.991
PLAOP4000	5	0.02178	2.8527	0.985
	10	0.08762	2.2913	0.992
	15	0.08904	2.6796	0.990

4. Conclusions

The addition of plasticizers and microfibers from OPF affected the thermal resistance properties of PLA biocomposites. PLAP4000 has better thermal resistance compared to PLAP200 and PLAG with T_{onset} and T_{max} values reaching 349.17°C and 374.68°C, respectively, approaching pure PLA. The addition of OPF reduces the thermal stability of all plasticized PLA, both with GTA and PEG. The crystallinity analysis of the DSC analysis showed that PEG has the ability as a nucleating agent better than GTA with the formation of an endothermic peak, a decrease in T_{cc} , and an increase in T_{c} in the exothermic process. All types of plasticizers reduced the T_{g} value by about 27–42%, while the addition of OPF reduced the T_{g} in PLA with the PEG plasticizer ranging from 37–55%. A review of crystallization kinetics shows that the crystallization rate increases with PEG and OPF plasticizers, which is around 4–9 times for PEG200, while 2–3 times for PEG4000. The crystal formation phenomenon occurs in 3-dimensional crystals with heterogeneous and simultaneous crystallization mechanisms where the value of n ranges from 2–3 at a cooling rate of 5–15°C/minute. PEG produces PLA composites that are better than GTA because it can function as a nucleating agent and a compatibilizer

between OPF and PLA. Overall, PEG4000 has better thermal resistance properties, a nucleating agent, and better compatibility than PEG200 and GTA, but the presence of OPF inhibits crystallization in PLA composites.

Acknowledgments

The author would like to thank the 2019 Thematic DIPA program from the Research center for Biomaterials - LIPI, which has funded this research activity. The author also thanks Dr. Lisman Suryanegara as the mentor in this activity, so that this research can run well.

References

- [1] M. S. Huda, L. T. Drzal, M. Misra, A. K. Mohanty, Wood-Fiber-Reinforced Poly(Lactic Acid) Composites: Evaluation of the Physicomechanical and Morphological Properties, *Journal of Applied Polymer Science*, 102, 5, (2006), 4856–4869 <https://doi.org/10.1002/app.24829>
- [2] Rahul M. Rasal, Amol V. Janorkar, Douglas E. Hirt, Poly(Lactic Acid) Modifications, *Progress in Polymer Science*, 35, 3, (2010), 338–356 <https://doi.org/10.1016/j.progpolymsci.2009.12.003>
- [3] Maria Laura Di Lorenzo, Crystallization Behavior of Poly(L-Lactic Acid), *European Polymer Journal*, 41, 3, (2005), 569–575 <https://doi.org/10.1016/j.eurpolymj.2004.10.020>
- [4] Sajjad Saeidlou, Michel A. Huneault, Hongbo Li, Chul B. Park, Poly(Lactic Acid) Crystallization, *Progress in Polymer Science*, 37, 12, (2012), 1657–1677 <https://doi.org/10.1016/j.progpolymsci.2012.07.005>
- [5] Rafael A. Auras, Bruce Harte, Susan Selke, Ruben Hernandez, Mechanical, Physical, and Barrier Properties of Poly(Lactide) Films, *Journal of Plastic Film & Sheeting*, 19, 2, (2003), 123–135 <https://doi.org/10.1177/8756087903039702>
- [6] O. Martin, L. Avérous, Poly(Lactic Acid): Plasticization and Properties of Biodegradable Multiphase Systems, *Polymer*, 42, 14, (2001), 6209–6219 [https://doi.org/10.1016/S0032-3861\(01\)00086-6](https://doi.org/10.1016/S0032-3861(01)00086-6)
- [7] B. Eling, S. Gogolewski, A. J. Pennings, Biodegradable Materials of Poly(L-Lactic Acid): 1. Melt-Spun and Solution-Spun Fibres, *Polymer*, 23, 11, (1982), 1587–1593 [https://doi.org/10.1016/0032-3861\(82\)90176-8](https://doi.org/10.1016/0032-3861(82)90176-8)
- [8] R. Vasanthakumari, A. J. Pennings, Crystallization Kinetics of Poly(L-Lactic Acid), *Polymer*, 24, 2, (1983), 175–178 [https://doi.org/10.1016/0032-3861\(83\)90129-5](https://doi.org/10.1016/0032-3861(83)90129-5)
- [9] W. Hoogsteen, A. R. Postema, A. J. Pennings, Gerrit Ten Brinke, P. Zugenmaier, Crystal Structure, Conformation and Morphology of Solution-Spun Poly(L-Lactide) Fibers, *Macromolecules*, 23, 2, (1990), 634–642 <https://doi.org/10.1021/ma00204a041>
- [10] L. T. Lim, R. Auras, M. Rubino, Processing Technologies for Poly(Lactic Acid), *Progress in Polymer Science*, 33, 8, (2008), 820–852 <https://doi.org/10.1016/j.progpolymsci.2008.05.004>
- [11] Nurul Hani Md Zubir, Sung Ting Sam, Nik Noriman Zulkepli, Mohd Firdaus Omar, The Effect of Rice

- Straw Particulate Loading and Polyethylene Glycol as Plasticizer on the Properties of Polylactic Acid/Polyhydroxybutyrate-Valerate Blends, *Polymer Bulletin*, 75, 1, (2018), 61-76
<https://doi.org/10.1007/s00289-017-2018-y>
- [12] Athanasia Amanda Septevani, Samsul Bhakri, Plasticization of Poly(Lactic Acid) Using Different Molecular Weight of Poly(Ethylene Glycol), *AIP Conference Proceedings*, 1904, 1, (2017), 020038
<https://doi.org/10.1063/1.5011895>
- [13] Bettina Dittrich, Karen-Alessa Wartig, Daniel Hofmann, Rolf Mülhaupt, Bernhard Schartel, The Influence of Layered, Spherical, and Tubular Carbon Nanomaterials' Concentration on the Flame Retardancy of Polypropylene, *Polymer Composites*, 36, 7, (2015), 1230-1241
<https://doi.org/10.1002/pc.23027>
- [14] Kyung-man Choi, Myeon-Cheon Choi, Dong-Hun Han, Tae-Sung Park, Chang-Sik Ha, Plasticization of Poly(Lactic Acid) (PLA) through Chemical Grafting of Poly(Ethylene Glycol) (PEG) Via in Situ Reactive Blending, *European Polymer Journal*, 49, 8, (2013), 2356-2364
<https://doi.org/10.1016/j.eurpolymj.2013.05.027>
- [15] Isabelle Pillin, Nicolas Montrelay, Yves Grohens, Thermo-Mechanical Characterization of Plasticized PLA: Is the Miscibility the Only Significant Factor?, *Polymer*, 47, 13, (2006), 4676-4682
<https://doi.org/10.1016/j.polymer.2006.04.013>
- [16] Justine Muller, Alberto Jiménez, Chelo González-Martínez, Amparo Chiralt, Influence of Plasticizers on Thermal Properties and Crystallization Behaviour of Poly(Lactic Acid) Films Obtained by Compression Moulding, *Polymer International*, 65, 8, (2016), 970-978
<https://doi.org/10.1002/pi.5142>
- [17] Buong Woei Chieng, Nor Azowa Ibrahim, Wan Md Zin Wan Yunus, Mohd Zobir Hussein, Plasticized Poly(Lactic Acid) with Low Molecular Weight Poly(Ethylene Glycol): Mechanical, Thermal, and Morphology Properties, *Journal of Applied Polymer Science*, 130, 6, (2013), 4576-4580
<https://doi.org/10.1002/app.39742>
- [18] Homero Salas-Papayanopolos, Ana B. Morales-Cepeda, Saúl Sanchez, Pierre G. Lafleur, I. Gomez, Synergistic Effect of Silver Nanoparticle Content on the Optical and Thermo-Mechanical Properties of Poly(L-Lactic Acid)/Glycerol Triacetate Blends, *Polymer Bulletin*, 74, 12, (2017), 4799-4814
<https://doi.org/10.1007/s00289-017-1992-4>
- [19] Xin-Feng Wei, Rui-Ying Bao, Zhi-Qiang Cao, Liang-Qing Zhang, Zheng-Ying Liu, Wei Yang, Bang-Hu Xie, Ming-Bo Yang, Greatly Accelerated Crystallization of Poly(Lactic Acid): Cooperative Effect of Stereocomplex Crystallites and Polyethylene Glycol, *Colloid and Polymer Science*, 292, 1, (2014), 163-172
<https://doi.org/10.1007/s00396-013-3067-x>
- [20] Xuetao Shi, Guangcheng Zhang, Thanh V. Phuong, Andrea Lazzeri, Synergistic Effects of Nucleating Agents and Plasticizers on the Crystallization Behavior of Poly(Lactic Acid), *Molecules*, 20, 1, (2015), 1579-1593
<https://doi.org/10.3390/molecules20011579>
- [21] Ali Abdulkhali, Jaber Hosseinzadeh, Saeed Dadashi, Mohammad Mousavi, A Study of Morphological, Thermal, Mechanical and Barrier Properties of PLA Based Biocomposites Prepared with Micro and Nano Sized Cellulosic Fibers, *Cellulose Chemistry and Technology*, 49, 7-8, (2015), 597-605
- [22] Yong-Qing Zhao, Hoi-Yan Cheung, Kin-Tak Lau, Cai-Ling Xu, Dan-Dan Zhao, Hu-Lin Li, Silkworm Silk/Poly(Lactic Acid) Biocomposites: Dynamic Mechanical, Thermal and Biodegradable Properties, *Polymer Degradation and Stability*, 95, 10, (2010), 1978-1987
<https://doi.org/10.1016/j.polyimdegstab.2010.07.015>
- [23] Nicolas Le Moigne, Marc Longerey, Jean-Marie Taulemesse, Jean-Charles Bénézet, Anne Bergeret, Study of the Interface in Natural Fibres Reinforced Poly(Lactic Acid) Biocomposites Modified by Optimized Organosilane Treatments, *Industrial Crops and Products*, 52, (2014), 481-494
<https://doi.org/10.1016/j.indcrop.2013.11.022>
- [24] Lisman Suryanegara, Antonio Norio Nakagaito, Hiroyuki Yano, Thermo-Mechanical Properties of Microfibrillated Cellulose-Reinforced Partially Crystallized PLA Composites, *Cellulose*, 17, 4, (2010), 771-778
<https://doi.org/10.1007/s10570-010-9419-5>
- [25] M. K. Mohamad Haafiz, Azman Hassan, Zainoha Zakaria, I. M. Inuwa, M. S. Islam, M. Jawaid, Properties of Polylactic Acid Composites Reinforced with Oil Palm Biomass Microcrystalline Cellulose, *Carbohydrate Polymers*, 98, 1, (2013), 139-145
<https://doi.org/10.1016/j.carbpol.2013.05.069>
- [26] E. W. Fischer, Hans J. Sterzel, G. Wegner, Investigation of the Structure of Solution Grown Crystals of Lactide Copolymers by Means of Chemical Reactions, *Kolloid-Zeitschrift und Zeitschrift für Polymere*, 251, 11, (1973), 980-990
<https://doi.org/10.1007/BF01498927>
- [27] Melvin Avrami, Kinetics of Phase Change. II Transformation-Time Relations for Random Distribution of Nuclei, *The Journal of Chemical Physics*, 8, 2, (1940), 212-224
<https://doi.org/10.1063/1.1750631>
- [28] Melvin Avrami, Kinetics of Phase Change. I General Theory, *The Journal of Chemical Physics*, 7, 12, (1939), 1103-1112
<https://doi.org/10.1063/1.1750380>
- [29] Mehmet Kodall, Humeyra Sirin, Guralp Ozkoc, Non-Isothermal Crystallization Kinetics of PEG Plasticized PLA/G-POSS Nanocomposites, *Polymer Composites*, 38, 7, (2017), 1378-1389
<https://doi.org/10.1002/pc.23704>
- [30] T. Zimmermann, E. Pöhler, T. Geiger, Cellulose Fibrils for Polymer Reinforcement, *Advanced Engineering Materials*, 6, 9, (2004), 754-761
<https://doi.org/10.1002/adem.200400097>
- [31] V. S. Giita Silverajah, Nor Azowa Ibrahim, Norhazlin Zainuddin, Wan Md Zin Wan Yunus, Hazimah Abu Hassan, Mechanical, Thermal and Morphological Properties of Poly(lactic acid)/Epoxidized Palm Olein Blend, *Molecules*, 17, 10, (2012), 11729-11747
<https://doi.org/10.3390/molecules171011729>
- [32] L. Quiles-Carrillo, S. Duart, N. Montanes, S. Torres-Giner, R. Balart, Enhancement of the Mechanical and Thermal Properties of Injection-Molded Polylactide Parts by the Addition of Acrylated Epoxidized Soybean Oil, *Materials & Design*, 140, (2018), 54-63
<https://doi.org/10.1016/j.matdes.2017.11.031>

- [33] Paola Giudicianni, Giuseppe Cardone, Raffaele Ragucci, Cellulose, Hemicellulose and Lignin Slow Steam Pyrolysis: Thermal Decomposition of Biomass Components Mixtures, *Journal of Analytical and Applied Pyrolysis*, 100, (2013), 213–222
<https://doi.org/10.1016/j.jaap.2012.12.026>
- [34] Sarifah Fauziah Syed Draman, Rusli Daik, Famiza Abdul Latif, Said M. El-Sheikh, Characterization and Thermal Decomposition Kinetics of Kapok (*Ceiba pentandra* L.)-Based Cellulose, *BioResources*, 9, 1, (2014), 8–23
- [35] Fatemeh Safdari, Pierre J. Carreau, Marie C. Heuzey, Musa R. Kamal, Effects of Poly(Ethylene Glycol) on the Morphology and Properties of Biocomposites Based on Polylactide and Cellulose Nanofibers, *Cellulose*, 24, 7, (2017), 2877–2893
<https://doi.org/10.1007/s10570-017-1327-5>
- [36] WeiDan Ding, Raymond K. M. Chu, Lun Howe Mark, Chul B. Park, Mohini Sain, Non-Isothermal Crystallization Behaviors of Poly(Lactic Acid)/Cellulose Nanofiber Composites in the Presence of CO₂, *European Polymer Journal*, 71, (2015), 231–247
<https://doi.org/10.1016/j.eurpolymj.2015.07.054>
- [37] S. Iannace, L. Nicolais, Isothermal Crystallization and Chain Mobility of Poly(L-Lactide), *Journal of Applied Polymer Science*, 64, 5, (1997), 911–919
[https://doi.org/10.1002/\(SICI\)1097-4628\(19970502\)64:5<911::AID-APP11>3.0.CO;2-W](https://doi.org/10.1002/(SICI)1097-4628(19970502)64:5<911::AID-APP11>3.0.CO;2-W)
- [38] Yong He, Zhongyong Fan, Yanfei Hu, Tong Wu, Jia Wei, Suming Li, DSC Analysis of Isothermal Melt-Crystallization, Glass Transition and Melting Behavior of Poly(L-Lactide) with Different Molecular Weights, *European Polymer Journal*, 43, 10, (2007), 4431–4439
<https://doi.org/10.1016/j.eurpolymj.2007.07.007>
- [39] Yonghui Li, Caihong Chen, Jun Li, Xiuzhi Susan Sun, Isothermal Crystallization and Melting Behaviors of Bionanocomposites from Poly(Lactic Acid) and TiO₂ Nanowires, *Journal of Applied Polymer Science*, 124, 4, (2012), 2968–2977
<https://doi.org/10.1002/app.35326>